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**Immediate action of SCN^- on the apical transport of
 Cl^- in the epithelial cells of rabbit gallbladder**

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Fisiologia. — *Immediate action of SCN^- on the apical transport of Cl^- in the epithelial cells of rabbit gallbladder (*)*. Nota di DARIO CREMASCHI, GIULIANO MEYER, CARLO ROSSETTI, GUIDO BOTTÀ e PAOLA PALESTINI, presentata (**) dal Corrisp. V. CAPRARO.

RIASSUNTO. — Il Tiocianato, posto nel lume della cistifellea di coniglio per 45 sec a varie concentrazioni (insieme col ^{36}Cl durante la misura del flusso dell'anione attraverso le membrane apicali e le giunzioni dell'epitelio), inibisce questo flusso di circa l'80% se usato in concentrazioni superiori a 17 mM circa. Riducendo i tempi di esposizione si ottiene lo stesso grado di inibizione. Si conclude che il Tiocianato opera legandosi ai siti apicali di trasporto del Cl^- inibendoli in modo pressoché immediato. Lo stiramento del tessuto aumenta la frazione insensibile al Tiocianato, mentre lascia invariata quella sensibile.

INTRODUCTION

In a previous paper we showed that SCN^- abolishes the apical entry of Cl^- into the epithelial cells of rabbit gallbladder [3]. The tissue was exposed to SCN^- for 45 min so that the doubt that the action could be indirect or metabolic was present, in spite of the fact that all results were consistent with a competition of SCN^- for Cl^- sites of transport.

The aim of this paper is to eliminate any doubt concerning this point by studying the action of the inhibitor on Cl^- influx through the apical membrane with very short times corresponding to those of the influx measurement (45 sec or less).

METHODS

New Zealand rabbits were killed by a blow on the neck. Gallbladders were excised and washed free from bile with Krebs-Henseleit solution (mM: 142.9 Na^+ , 127.7 Cl^- , 24.9 HCO_3^- , 5.9 K^+ , 1.2 $\text{H}_2\text{PO}_4^{2-}$, 1.2 SO_4^{2-} , 1.2 Mg^{2+} , 2.5 Ca^{2+}). Opened flat, they were mounted on a nylon mesh between two lucite chambers with the luminal surface facing upwards, exposed (0.61 cm^2) within the upper chamber, filled with 1 ml saline. After a pre-incubation period of 75 min, the tissue was bathed on the luminal side for 45 sec by the test so-

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lution added with ^3H -sucrose ($10\ \mu\text{Ci/ml}$) and ^{36}Cl ($4\ \mu\text{Ci/ml}$) in order to measure the Cl^- unidirectional influx into the epithelium through the apical membrane and the junctional complex. Test solutions were Krebs-Henseleit saline in which part of Cl^- was replaced by SO_4^{2-} and mannitol (control) or by SCN^- (experiment) at the reported concentrations. When SO_4^{2-} was used, mannitol was added to compensate osmolality. All salines were bubbled with $5\% \text{CO}_2 + 95\% \text{O}_2$; pH was 7.4 and temperature 27°C . Analyses were performed as reported in Ref. 2.

RESULTS

SCN^- effects. The action of SCN^- (present on the luminal side during the 45 sec measuring time at different concentrations) on Cl^- uptake (expressed as $\mu\text{eq cm}^{-2} \text{h}^{-1}$) is reported in fig. 1. It is to emphasize that maximal inhibition is obtained with concentrations larger than 17 mM SCN^- and that a residual Cl^- uptake, SCN^- independent (about $4\ \mu\text{eq cm}^{-2} \text{h}^{-1}$) is measured even under these conditions.

The action of the drug, used with a maximal concentration (25 mM), was also tested at different exposure times (30 and 45 sec). When a 30 sec period was employed, the flux measuring time was also reduced to this time (the mea-

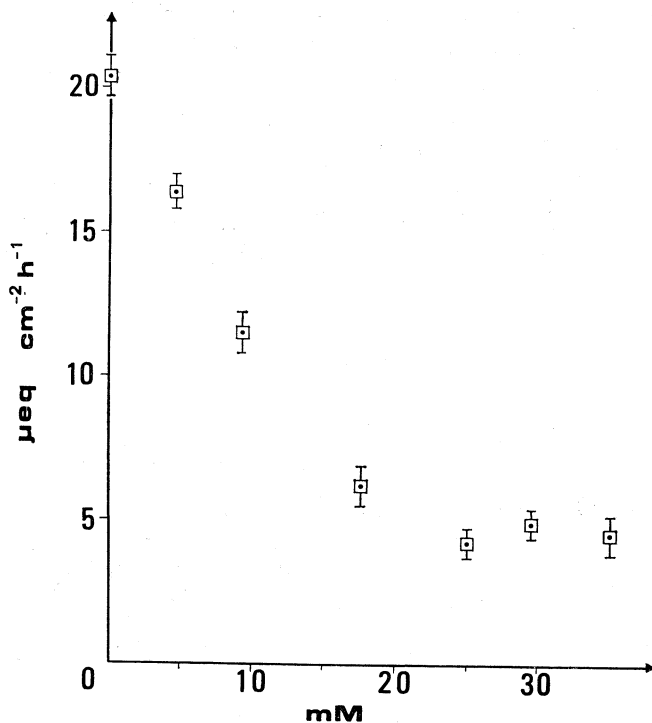


Fig. 1. - SCN^- effects on Cl^- influx. Abscissa: SCN^- concentrations (mM); Ordinata: Cl^- influx ($\mu\text{eq cm}^{-2} \text{h}^{-1}$). Results are reported as mean \pm standard error. (7 experiments per point).

sure of the unidirectional influx is constant if taken between 0 and 75 sec, see Ref. 3). The result is reported in Table I: the rate of Cl^- entry into the epithelium is inhibited to the same extent after both 30 and 45 sec of treatment.

Stretch effects. If the gallbladder, opened flat, is stretched during mounting, Cl^- influx increases significantly from 21.9 to 28.0 $\mu\text{eq cm}^{-2} \text{h}^{-1}$ (Table II).

TABLE I

Cl^- influxes (lumen to epithelium) at different times of measurement with different times of exposure to SNC^- .

In controls only $^{36}\text{Cl}^-$ was present in the luminal bath. In the experiments $^{36}\text{Cl}^-$ and SNC^- (25 mM) were simultaneously present in the luminal bath for 30 or 45 sec. Results are reported as means \pm standard error with the number of experiments in parenthesis.

P = statistical probability.

Exposure time (sec) to $^{36}\text{Cl}^-$ and SNC^-	Cl^- influx ($\mu\text{eq cm}^{-2} \text{h}^{-1}$)		P
	Control	SNC^-	
30	20.6 ± 2.5 (5)	4.4 ± 1.4 (6)	< 0.01
45	21.0 ± 1.2 (5)	4.0 ± 0.8 (6)	< 0.01
P	n. s.	n. s.	

TABLE II

SNC^- -dependent and SNC^- -independent Cl^- influxes (lumen to epithelium) measured under different stretch conditions of the tissue.

Results are reported as means \pm standard error with the number of experiments or freedom degrees (*fd*) in parenthesis. P = statistical probability.

	Unstretched conditions Cl^- influx ($\mu\text{eq cm}^{-2} \text{h}^{-1}$)	Stretched conditions Cl^- influx ($\mu\text{eq cm}^{-2} \text{h}^{-1}$)	P
Control	21.9 ± 1.1 (12)	28.0 ± 2.5 (12)	< 0.05
SNC^- present	4.6 ± 0.4 (23)	9.7 ± 1.4 (13)	< 0.01
Δ	17.3 ± 1.0 (<i>fd</i> = 33)	18.3 ± 2.8 (<i>fd</i> = 23)	n. s.

Correspondingly, in the presence of 25 mM SCN^- , the residual Cl^- influx is again significantly increased under the stretched condition (4.6 vs. 9.7 $\mu\text{eq cm}^{-2} \text{h}^{-1}$, see Table II), whereas the SCN^- -dependent fraction, calculated as a difference between the respective control and the residual Cl^- influx, is statistically equal (17.7 and 18.3 $\mu\text{eq cm}^{-2} \text{h}^{-1}$).

DISCUSSION

The fact that with 30 or 45 sec of exposure to SCN^- the same extent of inhibition is obtained points out that the maximal action of the drug is immediate or takes place with a lag negligible with respect to the measuring time (i.e. very few sec). This result, combined with the fact that the exposure to SCN^- is only luminal, confirms the previous [3] conclusion that the inhibitor acts directly on the transport sites of Cl^- , competing for them.

In the apical membrane no Cl^- conductance exists [4, 6] so that the entry of the anion is only neutral and mediated by a co-transport with Na^+ [2, 6]; thus, SCN^- should bind to the Cl^- sites of this co-transport. The fraction of Cl^- influx which turns out to be SCN^- -insensitive should be related to the slight Cl^- conductance of the junctional complex [1]. Stretch manoeuvres, which should affect paracellular pathways [1, 5], do not modify the SCN^- -sensitive component of Cl^- influx, whereas the SCN^- -insensitive fraction largely increases.

REFERENCES

- [1] BARRY P.H., DIAMOND J.M. and WRIGHT E.M. (1971) – *The mechanism of cation permeation in rabbit gallbladder. Dilution potentials and biionic potentials*. «J. Membrane Biol.», 4, 358–394.
- [2] CREMASCHI D. and HÉNIN S. (1975) – *Na^+ and Cl^- transepithelial routes in rabbit gallbladder. Tracer analysis of the transports*. «Pflügers Archiv-European J. Physiol.», 361, 33–41.
- [3] CREMASCHI D., HÉNIN S. and MEYER G. (1979) – *Stimulation by HCO_3^- of Na^+ transport in rabbit gallbladder*. «J. Membrane Biol.», 47, 145–170.
- [4] CREMASCHI D. and MEYER G. (1982) – *Amiloride-sensitive sodium channels in rabbit and guinea-pig gallbladder*. «J. Physiol.», 326, 21–34.
- [5] FRIZZEL R.A., DUGAS M.C. and SCHULTZ S.G. (1975) – *Sodium chloride transport by rabbit gallbladder. Direct evidence for a coupled NaCl influx process*. «J. Gen. Physiol.», 65, 769–795.
- [6] HÉNIN S. and CREMASCHI D. (1975) – *Transcellular ion route in rabbit gallbladder. Electric properties of the epithelial cells*. «Pflügers Archiv-European J. Physiol.», 355, 125–139.