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Radioimmunoassay of plasma estriol in males and females of Rana esculenta

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Biologia. — Radioimmunoassay of plasma estriol in males and females of Rana esculenta (*). Nota di Michela D'Istria, Franca Citarella e Giovanni Delrio, presentata (**) dal Socio G. Mon-Talenti.

RIASSUNTO. — La concentrazione plasmatica di estriolo in maschi e femmine di *Rana* esculenta è stata determinata col metodo radioimmunologico durante il ciclo annuale. L'estriolo è presente solo nelle femmine nel mese di dicembre (300 ± 105 pg/ml).

Estrogens have been measured in amphibian blood plasma by several Authors using the fluorimetric method or gas-chromatography (Gallien and Chalumeau Le Foulgoc, 1960; Cedard and Ozon, 1962; Polzonetti Magni *et al.*, 1970). d'Istria *et al.* (1974) performed the radioimmunoassay of testosterone, estradiol-17 β and estrone in the plasma of both sexes of *Rana esculenta* (Fig. 1) but did not assay estriol since a specific antiserum for this hormone was not easily available at that time.

The present report completes the annual cyclic pattern of plasma estrogens in *Rana esculenta* and in fact describes the determination of estriol by RIA in individual plasma samples from males and females during the sexual cycle; the method used was that of Youssefnejadian and Sommerville (1973).

Blood from 15 males and 15 females of Rana esculenta captured monthly in the surroundings of Naples was collected from the conus arterious in heparinized microtubes. H3-estriol (S.A. 85 Ci/mol.) from the Radiochemical Center Amersham (England) was used. The chemicals were analytical grade from Baker (U.S.A.). An estriol-6-carboximethyloxima-BSA antiserum, purchased from SORIN (Saluggia, Italy) was used. Aliquots of 200 µl or 400 µl of plasma were used for each assay. The plasma plus 2000 dpm of H³-estriol was extracted twice with 10 ml of diethyl ether; the extracts, transferred to a pointed tube, were dried at 50 °C under a stream of nitrogen. The residues were removed with four drops of benzene-methanol 85:15 (v/v) containing a yellow dye isatin (0.1 %) and transferred to a column (Pasteur pipette) of Sephadex LH 20 equilibrated with benzene-methanol 85:15 (v/v). Elution was performed with the same solvents. The yellow fraction containing estradiol was discarded and immediately a fraction of 4 ml was collected from the column in a counting vial, dried under nitrogen at 40 °C and redissolved in acetone. Appropriate aliquots were removed for assay and the remainder was subjected to liquid scintillation to determine the recovery.

(**) Nella seduta dell'11 marzo 1978.

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Standard curve (10–200 pg) and unknown samples were prepared in duplicate, dried under nitrogen at 50 °C, cooled to room temperature and equilibrated with 100 μ l of diluted antiserum (1/10,000) in phosphate buffer pH 7 for 10'. After addition of 100 μ l of H³-estriol (20,000 dpm) in phosphate



Fig. 1. – Plasma levels of testosterone, estradiol-17 β and estrone in the female of *Rana esculenta*. The arrow indicates the estriol presence in December (300 pg/ml); this hormone was not detectable in other periods of the year. $E_1 =$ estrone; $E_2 =$ estradiol-17 β ; $E_3 =$ Estriol test = testosterone.

buffer, the solutions were mixed and equilibrated at 4 °C overnight. The free steroid was removed by the addition of dextran-coated charcoal. The accuracy, precision, sensitivity and specificity were controlled using amphibian plasma. Specific quantities of estriol were added to samples containing a known amount of hormone. The range of estriol added was 80-1500 pg and the recovery % (mean \pm S.D.) was 95 ± 9 . The lower limit of sensitivity was 5 pg, whilst the precision (coefficient of variation % limits) was 9.4-14.

The antiserum had a cross reaction of less than 0.0005 % with testosterone, progesterone, corticosterone, cortisol, cholesterol and estrone, and less than 0.005 % with estradiol-17 β .

Estriol was never detected in male plasma. In females it was detectable just in December (13 animals out of 15) with a mean value of 300 ± 105 pg/ml, whereas in other periods of the year the amount is below the sensitivity of the method.

At this juncture the data obtained on the same species, using a gas chromatographic method by Polzonetti *et al.* (1970) must be brought into discussion because there is some perplexity as to the plasma value of estriol (110–360 ng/ml) reported by these Authors. This is a very high amount of estriol and, besides, it is strange that in the ovarian tissue extract we never detected the presence of this hormone using gas-chromatographic analysis (unpublished data). The value obtained by RIA is more credible, especially if compared with that obtained for estriol-17 β and estrone in the plasma of the same species.

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